Serum S100 β as a predictor of severity and outcomes for mixed subtype acute ischaemic stroke

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INTRODUCTION Serum S100 β levels are mostly used for predicting outcomes of large-vessel stroke. Its application to mixed subtypes of acute ischaemic stroke (AIS) has been limited.

METHODS Patients with mixed subtypes of AIS who were aged over 18 years and presented within 24 hours of stroke onset were consecutively enrolled. Serum S100 β levels at presentation (S100 β_b) and 72 hours (S100 β_{72hrs}), and corresponding National Institutes of Health Stroke Scale (NIHSS_b and NIHSS_{72hrs}, respectively) scores were assessed. Stroke outcomes were evaluated using the modified Rankin Scale (mRs) at 30 days (mRs₃₀) and 90 days (mRs₉₀). Correlations between S100 β_{patrs} , as well as differences between the two (Δ S100 β) and the corresponding NIHSS, mRs₃₀ and mRs₉₀ scores, were evaluated (p < 0.05).

RESULTS 35 patients were eligible for analysis. On univariate analysis, stroke outcomes had a significant association with $S100\beta_{b}$, $S100\beta_{72hrs}$, $NIHSS_{b}$, $NIHSS_{72hrs}$ and $\Delta S100\beta$. Both $S100\beta_{b}$ and $S100\beta_{72hrs}$ correlated with corresponding NIHSS values (ρ_{b} = 0.51, p < 0.001; ρ_{72hrs} = 0.74, p < 0.001), mRs₃₀ (ρ_{b} = 0.58, p < 0.001; ρ_{72hrs} = 0.72, p < 0.001) and mRs₉₀ (ρ_{b} = 0.51, p = 0.002; ρ_{72hrs} = 0.68, p < 0.001). Correlations existed between Δ S100 β and mRs₃₀ (ρ = 0.74, p < 0.001) and mRs₉₀ (ρ = 0.71, p < 0.001). Practical cut-off points for unfavourable outcomes (mRs 3–6) were S100 β_{72hrs} > 0.288 µg/L (sensitivity 92.3%, specificity 86.4%) and Δ S100 β > 0.125 µg/L (sensitivity 100%, specificity 81.8%).

CONCLUSION High serum S100 β is associated with unfavourable outcomes for mixed subtype AIS. Cut-off values of S100 β_{72hrs} and Δ S100 β were optimal for predicting unfavourable stroke outcomes.

Keywords: acute ischaemic stroke, outcome, serum S100ß

INTRODUCTION

Various neural-specific biomarkers are released into the serum and cerebrospinal fluid during neural tissue injuries. Both direct and indirect cerebral damage (e.g. cerebral trauma, cardiopulmonary arrest, postoperative neurovascular complications and acute ischaemic stroke [AIS]) lead to the release of these biomarkers.⁽¹⁻⁵⁾ Thus, assessment of the presence and magnitude of such biomarkers during neurovascular insults may facilitate not only the diagnosis but also the prediction of the complications and final outcomes of the neural injury.^(1,2,6) A recent study found that each biomarker has a specific cellular origin within the cerebral tissue (e.g. neuron-specific enolase [NSE] is released from neurons; serum S100^β is released from astrocytes and Schwann cells; Tau protein is from neurons; metalloproteinase-9 is released from vascular endothelium; and ubiquitin C terminal hydroxylase-L1 is released from neurons).⁽³⁾ Therefore, several biomarkers associated with specific cerebral cellular damage have been under extensive investigation to identify novel biomarkers that could serve these purposes.⁽⁷⁾ Recently, among many stroke-related biomarkers that were widely evaluated for their suitability for clinical deployment, S100β and NSE were the two most broadly attributed in many clinical studies for their clinical usefulness.(2,3,6,8-12) Because of the greater clinically correlated response of S100^β and absence of interference by the body's clearance systems, there has been much interest in proving its clinical applicability.⁽³⁾

The S100β protein is an acidic, calcium-binding, neuralspecific biomarker. It is a ~21-kDa protein with a homodimeric structure (double helix loop) that is encoded on the long arm of chromosome 21 (21q22.3).^(2,13) As S100β has been used as a target of immunologically specific staining techniques to demonstrate astrocytes and melanocytes, it is a potentially promising biomarker for damage to astroglial cells as well as melanocyte proliferation.^(3,7) Furthermore, a study based on experimental induction of endothelial injury has suggested that S100β could be a biomarker for blood-brain barrier disruption.⁽¹⁴⁾

Several studies have reported a significant elevation of serial S100 β levels following AIS, in particular among patients with large-vessel or cortical brain infarction. The elevation of S100 β , both initially and subsequently at the appropriate time points, significantly correlated with not only the extent of cerebral infarction, as reflected by stroke severity, but also the associated complications and neurological outcomes.^(B-12,15,16)

Earlier studies have reported that S100 β was first detected in the serum of patients with AIS at 6–12 hours and reached peak levels at 72 hours after the onset of AIS.^(12,17,18) It followed firstorder kinetics, with a biological half-life of 25.3 ± 5.1 minutes, without effect from a moderately decreased glomerular filtration rate.⁽⁵⁾ Nevertheless, due to the delayed kinetics for the first detection in serum (6–12 hours), S100 β is not an ideal biomarker for AIS diagnosis.^(12,18)

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To the best of our knowledge, there has been limited evaluation of the clinical usefulness of S100 β in mixed cortical (or large-vessel occlusive stroke [LVS]) and subcortical (or small-vessel occlusive stroke [SVS]) AIS. Therefore, in the present study, we aimed to: (a) demonstrate the significance of the correlations between the initial and subsequent (at 72 hours after AIS onset) S100 β levels with the severity and outcomes of patients with mixed subtype AIS; and (b) determine the optimal cut-off points of S100 β levels at each time point for predicting unfavourable outcomes in patients with mixed subtype AIS. We intended to prove the clinical applicability of S100 β for predicting the outcomes of patients with mixed subtype AIS.

METHODS

We prospectively enrolled all patients aged over 18 years who presented to Songklanagarind Hospital, Faculty of Medicine, Prince of Songkla University, Songkhla, Thailand, from January 2013 to September 2013 with newly diagnosed AIS within 24 hours of the onset of symptoms. The diagnosis of AIS was based on the patient's clinical presentation and confirmed by either computed tomography or magnetic resonance imaging of the brain. Patients with a previous history of neurological illness, existing neurological disorder or residual neurological disability from any cause (e.g. stroke, cerebral neoplasm, intracranial infection or traumatic cerebral injury) were excluded. To achieve statistically significant power, a sample size of 36 was necessary based on the equation:

$$\therefore n_{l} = \frac{\left[Z_{1-\alpha/2}\sqrt{p(1-p)(1+1/r)} + Z_{\beta}\sqrt{\frac{p(1-p)(1+p)}{p^{2}(1-p^{2})/r}}\right]}{\Delta^{2}}$$

where proportions of positive outcomes among the exposed group (p1) = 0.73 and proportions of positives outcome among the nonexposed group (p2) = 0.17, $\alpha = 0.05$, power = 0.8, and r = 1.

Patients' demographic data and cardiovascular risk factors were collected during stroke risk screening. Presenting stroke severity was graded using the National Institutes of Health Stroke Scale score (NIHSS) immediately upon the patient's arrival at our centre, as the baseline severity scale (NIHSS_b). An emergency cranial imaging study, in most cases computed tomography of the brain, was performed to confirm the diagnosis and classify the subtype of AIS, in combination with the clinical stroke syndrome. All brain images were subsequently reviewed and confirmed by a clinically blinded neuroradiologist. The first blood sample for the baseline assay of S100 β (S100 β _L) was collected prior to starting treatment.

The primary AIS treatment consisted of one or a combination of the following: intravenous thrombolysis, endovascular thrombectomy and antithrombotic agents. The second S100β assay was performed at 72 hours after the onset of stroke symptoms (S100 β_{72hrs}), together with a re-evaluation of stroke severity using NIHSS (NIHSS_{72hrs}). Stroke-related complications (e.g. haemorrhagic transformation and malignant cerebral oedema necessitating decompressive craniectomy) were monitored. Stroke outcomes were assessed using the modified Rankin Scale (mRs) at 30 days (mRs₃₀) and 90 days (mRs₉₀) after AIS onset. mRs ≤ 2 was considered as favourable, while mRs 3–6 was unfavourable. Measurement of serum S100 β levels was performed using a commercial kit (Elecsys S100; Roche Diagnostics, Indianapolis, IN, USA) that was composed of a two-site monoclonal S100-specific antibody to form a sandwich complex on a fully automated system (Elecsys 2010, Modular Analytics E170; Roche Diagnostics-Hitachi High-Technologies Corporation, Tokyo, Japan). The range of measurement was 0.015–30.0 µg/L. Values were evaluated within an intra-assay coefficient of variation in the range of 1.28%–2.32%.

Descriptive statistics were shown as number and percentage, mean ± standard deviation or median (interguartile range [IQR]). Fisher's exact test and chi-square test were used to analyse categorical variables, whereas Student's t-test and Wilcoxon ranksum test were used to analyse continuous variables for statistical significance. A p-value < 0.05 was considered to be statistically significant. Continuous variables were assessed using the Shapiro-Wilk test for normality. Correlations between S100B₆ and S100B_{73ber} and the corresponding NIHSS_b and NIHSS_{72brs} values as well as correlations between each S100ß level and the difference of the two S100 β measurements (Δ S100 β) with mRs₃₀ and mRs₉₀ were evaluated using Spearman's correlation. The optimal cut-off points of $S100\beta_{b'}$ $S100\beta_{72bs}$ and $\Delta S100\beta$ levels to predict final patient outcomes and mortality were obtained using receiver operating characteristic (ROC) curves. The sensitivity, specificity, positive predictive value and negative predictive value of each S100ß and Δ S100 β cut-off point obtained were also calculated.

The complete study protocol was reviewed and approved by the ethics committee of the Faculty of Medicine, Prince of Songkla University, Songkhla, Thailand. We strictly followed the regulations of the 1964 Declaration of Helsinki and its later amendments while performing the study. All identifiable personal information of enrolled patients was made completely anonymous.

RESULTS

In total, 35 out of 44 enrolled patients with AIS were eligible for final analysis after excluding six patients who met the exclusion criteria and three patients who had incomplete follow-up. There were 26 men and nine women with an overall mean age of 65.3 ± 10.7 (range 43–81) years (Table I). The men were slightly older than the women.

Smoking, hypertension and hyperlipidaemia were sequentially the three most common cardiovascular risk factors among our patients. The total number of SVSs was 17 (48.6%), involving 15 SVSs and two other undetermined subtypes that were finally classified as SVSs. The median NIHSS_b was 10.0 (IQR 6.0–19.0), while the median NIHSS_{72hrs} was 6.0 (IQR 2.5–16.0). The S100 β_b assessments were available at a mean time of 10.6 ± 7.3 (range 2.0–24.0) hours after stroke onset, while the S100 β_{72hrs} assessments were performed at 72 hours after stroke onset.

For primary stroke treatment, 21 patients received conventional therapy in the form of antiplatelet or anticoagulant therapy, while ten patients received intravenous thrombolysis and one patient underwent endovascular thrombectomy. Primary stroke treatment was withheld for three patients – two patients had haemorrhagic transformation and one patient had massive cerebral oedema

Table I. Univariate logistic regression analysis of clinical variables and stroke outcomes at 30 days and 90 days.

Variable			p-value		
	Total (n = 35)	mRs 0–2* (n = 22)	mRs 3–6* (n = 13)		
Gender				> 0.999	
Male	26 (74.3)	16 (72.7)	10 (76.9)		
Female	9 (25.7)	6 (27.3)	3 (23.1)		
Age ⁺ (yr)	65.3 ± 10.7	63.8 ± 10.2	67.8 ± 11.5	0.289	
Male	66.5 ± 11.1				
Female	61.6 ± 8.9				
Risk factor					
Smoking	20 (57.1)	12 (54.5)	8 (61.5)	0.960	
Hypertension	18 (51.4)	12 (54.5)	6 (46.2)	0.897	
Dyslipidaemia	10 (28.6)	8 (36.4)	2 (15.4)	0.259	
Diabetes mellitus	7 (20.0)	2 (9.1)	5 (38.5)	0.075	
Atrial fibrillation	6 (17.1)	2 (9.1)	4 (30.8)	0.166	
Coronary artery disease	4 (11.4)	1 (4.5)	3 (23.1)	0.134	
TOAST classification					
Large vessel	13 (37.1)	5 (22.7)	8 (61.5)		
Small vessel	15 (42.9)	14 (63.6)	1 (7.7)		
Cardioembolic	5 (14.3)	1 (4.5)	4 (30.8)		
Others (determined)	0 (0)	0 (0)	0 (0)		
Others (undetermined)	2 (5.7)	2 (9.1)	0 (0)		
NIHSS [‡]					
At baseline	10.0 (6.0–19.0)	7.0 (4.2–9.8)	20.0 (19.0–21.0)	< 0.001§	
At 72 hr	6.0 (2.5–16.0)	3.0 (2.0–4.8)	18.0 (16.0–28.0)	< 0.001§	
S100β [‡] (μg/L)					
At baseline	0.1 (0.1–0.2)	0.1 (0.1–0.1)	0.2 (0.1–0.2)	< 0.001§	
At 72 hr	0.2 (0.1–0.7)	0.1 (0.1–0.2)	1.0 (0.5–4.1)	< 0.001§	
Difference from baseline	0.1 (0.0–0.5)	0 (0.0–0.1)	0.9 (0.3–3.8)	< 0.001§	
Treatment					
Conventional therapy	21 (60.0)	16 (72.7)	5 (38.5)		
Intravenous thrombolysis	10 (28.6)	5 (22.7)	5 (38.5)		
Endovascular thrombectomy	1 (2.9)	0 (0.0)	1 (7.7)		
Haemorrhagic transformation	5 (14.3)	1 (4.5)	4 (30.8)		
Decompression craniotomy	2 (5.7)	0 (0.0)	2 (15.4)		

*mRs score at 30 days and 90 days after stroke were identical (mRs 0-2 = favourable outcome, mRs 3-6 = unfavourable outcome). †Data presented as mean ± standard deviation. ‡Data presented as median (interquartile range). Sp < 0.05 was statistically significant using Mann-Whitney *U* test. mRs: modified Rankin Scale score at 30 days and 90 days after stroke; NIHSS: National Institutes of Health Stroke Scale score; S100 β : serum S100 β ; TOAST classification: Trial of ORG 10172 in acute stroke treatment classification

initially. The final stroke outcomes evaluated using mRs₃₀ and mRs₉₀, which were categorised as favourable (mRs \leq 2) or unfavourable (mRs 3–6), were identical for the categorical outcomes (favourable or unfavourable) at both time points for each patient. Five patients had complications of haemorrhagic transformation, and two patients developed malignant brain oedema for which decompression craniotomy was indicated. Eventually, 22 patients had a favourable outcome, while 13 patients had an unfavourable outcome. Six of the patients died of direct stroke-related severity and complications.

 $S100\beta_{b'}~S100\beta_{72hrs'}~\Delta S100\beta,~NIHSS_b$ and $NIHSS_{72hrs}$ were significantly associated with categorical stroke outcomes on

univariate logistic regression analysis (Table I). Further evaluation of S100 β_b showed positive correlations with NIHSS_b ($\rho = 0.51$, p < 0.001), mRs₃₀ ($\rho = 0.58$, p < 0.001) and mRs₉₀ ($\rho = 0.51$, p = 0.002) using Spearman's correlation (Fig. 1). S100 β_{72hrs} also had positive correlations with NIHSS_{72hrs} ($\rho = 0.74$, p < 0.001), mRs₃₀ ($\rho = 0.72$, p < 0.001) and mRs₉₀ ($\rho = 0.68$, p < 0.001) (Fig. 2). The difference between S100 β_{72hrs} and S100 β_b (i.e. Δ S100 β) also showed a positive correlation with mRs₃₀ ($\rho = 0.74$, p < 0.001) and mRs₉₀ ($\rho = 0.74$, p < 0.001) and mRs₉₀ ($\rho = 0.71$, p < 0.001) but no significant correlation with the difference between NIHSS_{72hrs} and NIHSS_b (i.e. Δ NIHSS, $\rho = 0.07$, p = 0.700) (Fig. 3).

The optimal cut-off points for $S100\beta_{b'} S100\beta_{72hrs}$ and $\Delta S100\beta$ for the prediction of unfavourable stroke outcomes and death,



Fig. 1 Charts show the correlation of serum S100 β at baseline with (a) NIHSS score at baseline; (b) mRs score at 30 days after stroke; and (c) mRs score at 90 days after stroke. mRs: modified Rankin Scale; NIHSS_b: National Institute of Health and Stroke Scale score



Fig. 2 Charts show correlation of serum S100 β at 72 hours after stroke onset with (a) NIHSS score at 72 hours after stroke onset; (b) mRs score at 30 days after stroke; and (c) mRs score at 90 days after stroke. mRs: modified Rankin Scale; NIHSS: National Institutes of Health Stroke Scale score



Fig. 3 Charts show correlation of Δ S100 β with (a) Δ NIHSS; (b) mRs score at 30 days after stroke; and (c) mRs score at 90 days after stroke. Δ NIHSS: difference between National Institutes of Health Stroke Scale score at baseline and at 72 hours after stroke onset; Δ S100 β : difference between serum S100 β at baseline and at 72 hours after stroke onset; mRs: modified Rankin Scale

as well as sensitivity, specificity, area under the curve, positive predictive value and negative predictive value identified by the ROC curves, are presented in Table II.

DISCUSSION

Unlike most previous studies that involved large vessel thrombotic and embolic strokes, 17 (48.6%) of 35 patients in our study were

classified as SVS. We found that $S100\beta_b$, $S100\beta_{72hrs'}$ $\Delta S100\beta$, NIHSS_b and NIHSS_{72hrs} had a significant association with final stroke outcomes upon univariate logistic regression analysis. However, statistical significance was not found following multivariate logistic regression analysis of all NIHSS and S100 β values. Therefore, S100 β_b and S100 β_{72hrs} may not be superior to NIHSS_b and NIHSS_{72hrs} for predicting the final functional outcome of patients with stroke.

Prediction	S100β level (μg/L)	Sensitivity (%)	Specificity (%)	AUC	PPV (%)	NPV (%)
Unfavourable outcome (n = 13)						
All stroke (n = 13)						
<i>S100β_b</i>	0.096	92.3	68.2	0.80	63.0	93.0
S100B _{72hrs}	0.288	92.3	86.4	0.89	80.0	95.0
ΔS100β	0.125	100.0	81.8	0.91	76.5	100.0
Large vessel stroke (n = 12)						
S100β _b	0.158	75.0	100.0	0.88	100.0	67.0
S100B _{72hrs}	0.745	75.0	100.0	0.88	100.0	67.0
ΔS100β	0.294	83.3	83.3	0.83	90.9	71.4
Small vessel stroke $(n = 1)$						
<i>S100β</i> _b	0.098	100.0	69.0	0.84	16.0	100.0
S100 β_{72hrs}	0.242	100.0	87.0	0.94	33.0	100.0
Δ5100β	0.125	100.0	81.2	0.90	25.0	100.0
Death (n = 6)						
At 30 days						
S100β _b	0.158	100.0	75.0	0.88	27.3	100.0
S100 β_{72hrs}	0.375	100.0	65.6	0.83	21.4	100.0
At 90 days						
<i>S100β_b</i>	0.096	100.0	48.3	0.74	28.6	100.0
S100β _{z2hrs}	0.242	100.0	62.1	0.81	35.3	100.0

Table II. Optimal cut-off points for serum S100β at baseline and at 72 hours after stroke onset for predicting unfavourable stroke outcome (mRs > 2) and death (mRs 6) at 30 days and 90 days after stroke.

ΔS100β: difference between serum S100β at baseline and at 72 hours after stroke onset; AUC: area under the curve; mRs: modified Rankin Scale; NPV: negative predictive value; PPV: positive predictive value; S100β_{22hg}: serum S100β at 72 hours after stroke onset; S100β₂; serum S100β at baseline

Several widely reported previous clinical studies regarding the usefulness of \$100ß and other more specific neuronal biomarkers were based on patients with LVS and mostly middle cerebral artery occlusion. Foerch et al⁽¹⁶⁾ reported a correlation between a single S100ß measurement and functional outcome and also brain infarction volume seven days after stroke. They also found that the S100ß level was not influenced by variables such as age, gender, stroke severity, aetiology, size of lesion and risk factors upon multivariate logistic regression analysis.⁽¹⁶⁾ Furthermore, a single-measurement study of S100^β at 48 hours after stroke onset reported that S100 β level \leq 0.37 µg/L could predict an independent functional outcome, with a sensitivity of 87% and specificity of 78% for patients with middle cerebral artery infarction.⁽¹⁶⁾ Another study confirmed that a S100ß cut-off level > 0.2 μ g/L at 48 hours after stroke in patients with AIS was a strong predictor for unfavourable functional outcome at three months after stroke onset, with a sensitivity of 85% and specificity of 92%. Moreover, a study demonstrated that elevation of \$100ß as early as six hours after stroke onset was associated with an unfavourable functional outcome.⁽¹²⁾ Fassbender et al⁽⁸⁾ showed a correlation between elevated serum S100^β levels at ten hours, 24 hours and 72 hours after stroke onset with an infarction volume > 5 mm³ and functional outcome. In their study, the elevation of S100^β was found to be significantly higher for cortical infarctions than for subcortical and brain stem infarctions at 24 hours and 72 hours after stroke onset.(8)

In our study, the first blood sample for the S100 β assay was collected within 12 hours and, significantly, a second sample was

collected at 72 hours after stroke onset, which was considered the most optimal time point to assess S100 β levels for estimating the extent of cerebral damage after stroke. This time point for the second S100 β measurement was selected based on the findings of a previous experimental animal study and a clinical review of the role of S100 β in AIS.^(17,19) Although the level of S100 β in our study was not identical to previous studies, we found a comparable correlation with stroke severity as well as short- and long-term functional outcomes, as evaluated by mRs₃₀ and mRs₉₀, among our patients. Similarly, Δ S100 β , in our study, showed a correlation with mRs₃₀ and mRs₉₀.

The cut-off values of S100 β_b , S100 β_{72hrs} and Δ S100 β were good predictors of unfavourable outcome for all stroke subtypes in our study. However, as only one patient with SVS who experienced unfavourable outcome was included, our findings vis-à-vis the preciseness and generalisation of the cut-off values for S100 β to all patients with stroke may be limited. Additionally, the cut-off values found for S100 β_{72hrs} and Δ S100 β were notably more appropriate than S100 β_b in predicting an unfavourable functional outcome. Therefore, similar to previous studies,^(8,17,19) we also propose that S100 β measurement is appropriate for the prediction of unfavourable stroke outcome among patients with LVS, while its usefulness in patients with SVS remains as yet undetermined. Furthermore, a single measurement at 72 hours, or S100 β_{72hrs} , has the highest specificity for predicting an unfavourable outcome of AIS.

Most studies have recognised the clinical usefulness of assessments of S100 β levels for estimating the extent of cerebral

ischaemic damage that is reflected in infarction volume, monitoring patients' response to treatment and their subsequent functional outcome, as assessed by mRs.^(6,8,12,15,16,18,20) The S100 β level can be detected in the serum as early as 6–12 hours after stroke onset and, in most patients, reaches its peak 24 hours or more after stroke.⁽¹⁸⁾ However, some other studies have reported the S100B peak at 48-72 hours after stroke onset,^(8,9,12,17) suggesting that there is as yet no consensus over the optimal time points or number of \$100^{\beta} sample assays to be rationally performed for patients with stroke. On the contrary, certain studies have suggested that cerebrospinal fluid might be a more appropriate source for S100β and NSE assessments to predict stroke severity since, with cerebrospinal fluid measurements, there is no interference from other potential extracranial sources of \$100ß such as adipose tissue, melanocytes, T-lymphocytes, skin or skeletal muscles.(2,11) In view of these earlier findings, physicians should exercise caution when using \$100\beta and NSE measurements for evaluating patients with stroke unless such potential extracranial sources are carefully excluded.

Other biomarkers for the prediction of stroke outcome have also been investigated. Jauch et al, in the National Institute of Neurological Disorders and Stroke Recombinant Tissue Plasminogen Activator Stroke Study, demonstrated a positive association between the peak 24-hour level of myelin basic proteins, NSE, S100 β and NIHSS at presentation, as well as a worse outcome at three months after stroke among patients with a higher difference of S100 β and myelin basic proteins levels between 24 hours and two hours after initiation of intravenous thrombolysis treatment.⁽⁶⁾ Although NSE followed the same kinetic pattern as S100 β , it showed no correlation with infarction volume and functional outcomes,⁽⁸⁾ suggesting that NSE has no clinical efficacy for the prediction of outcome of AIS.

A difference in the elevation of biomarkers among patients with LVS and SVS has not been clearly elucidated.⁽²⁾ A study on glial tissue-specific proteins suggested that glial fibrillary acidic protein was more sensitive for estimating brain damage in patients with SVS, whereas S100 β was more suitable for patients with LVS.⁽¹¹⁾

The present study, which was conducted in Thailand, demonstrated that $S100\beta$ had a high specificity for predicting unfavourable outcomes of LVS in an Asian population. Differences in ethnicity have no influence on the S100 β measurements. Further clinical investigations enrolling a larger sample size of various stroke subtypes, with particular emphasis on patients with SVS, are warranted to confirm the true clinical usefulness of S100 β as a broad stroke biomarker. The optimal number of assessments required and the appropriate time points at which to perform these S100 β assessments also need to be specified to standardise practices and

to gain a better understanding of the diagnostic and prognostic value of $$100\beta$ measurements among patients with stroke.

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